



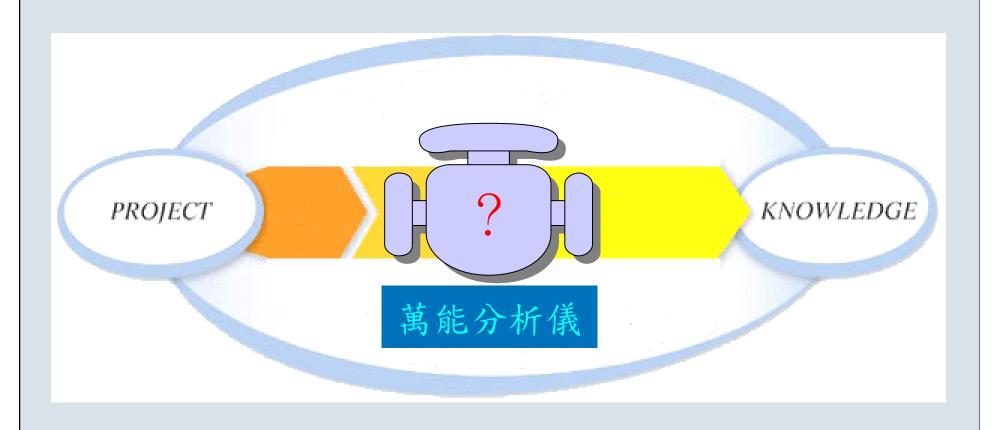
# The skill of LC-ESI-Ion Trap

For Proteomics Research

**James Yu, Application Specialist** 

# 科學家的共同夢想





**Bruker Daltonics** 

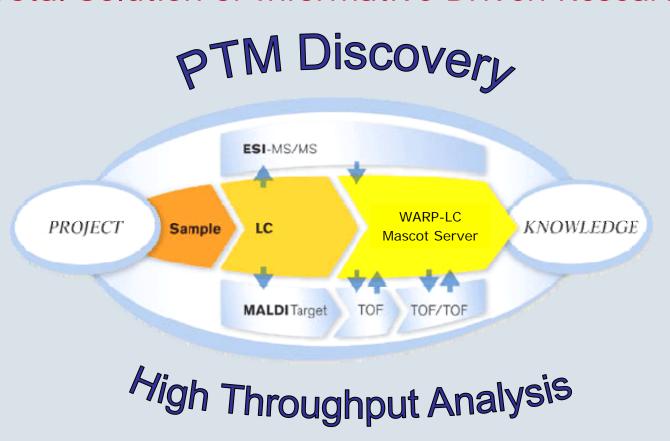


# 世事總難盡如人意

### Proteineer LC TM Workflow



Total Solution of Informative Driven Research



### What Do We See?

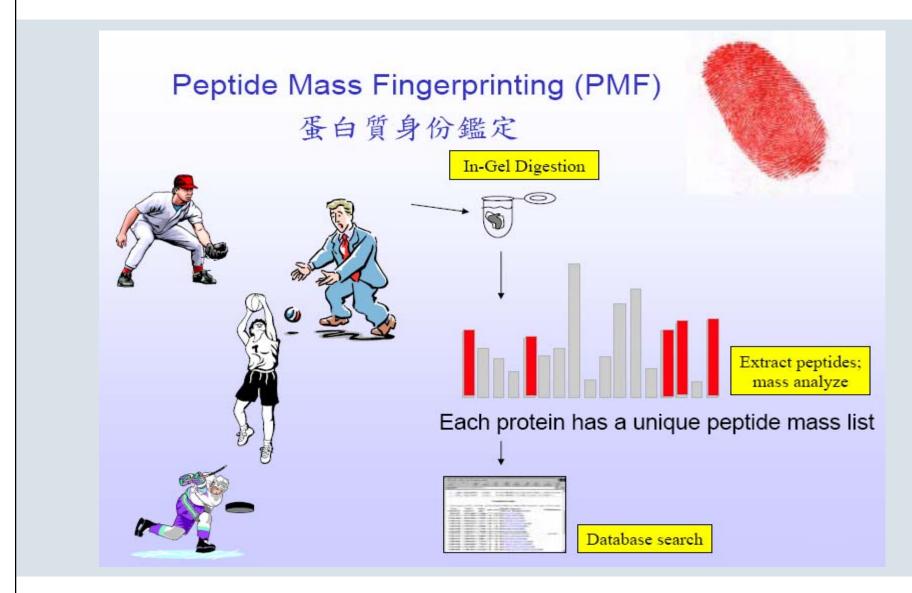


Is



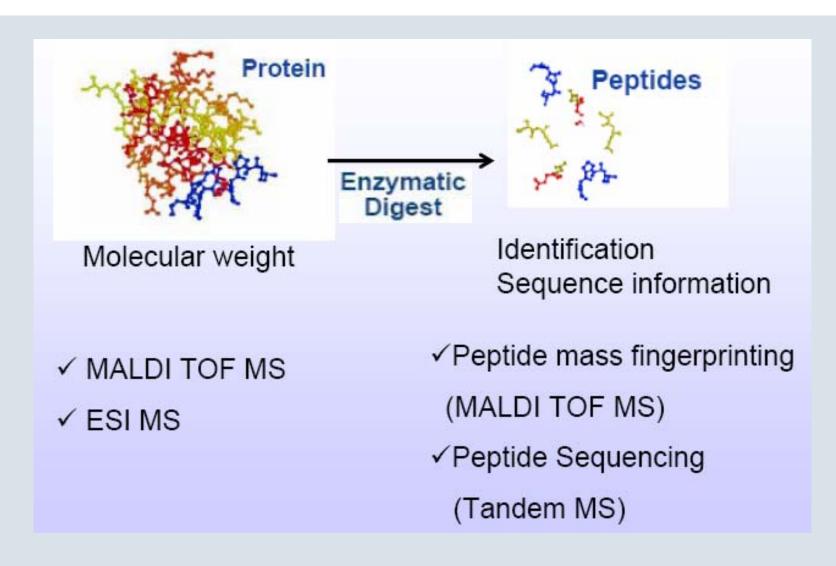
### What's the different between these people?





### **Mass Spectrometry Methods**

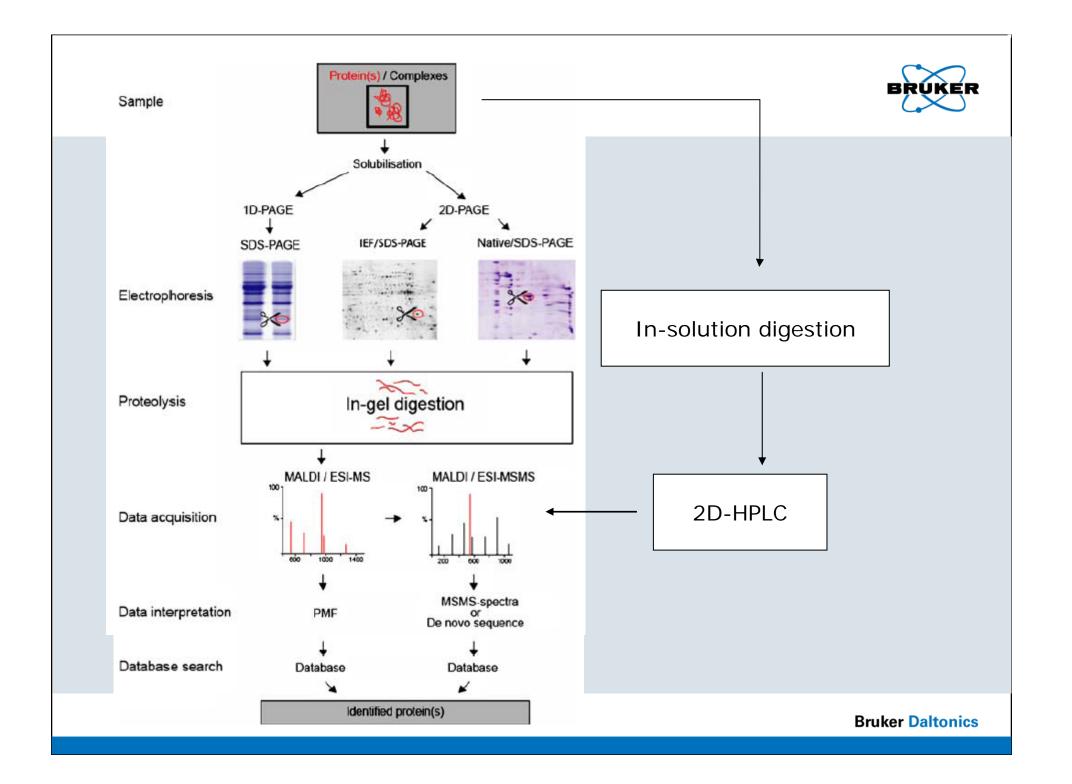




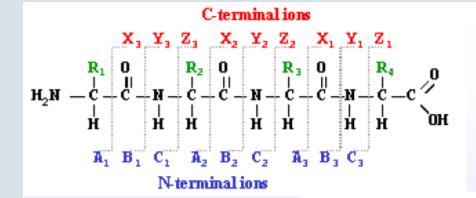
### What's the different between these people?











# Peptide Fragmentation

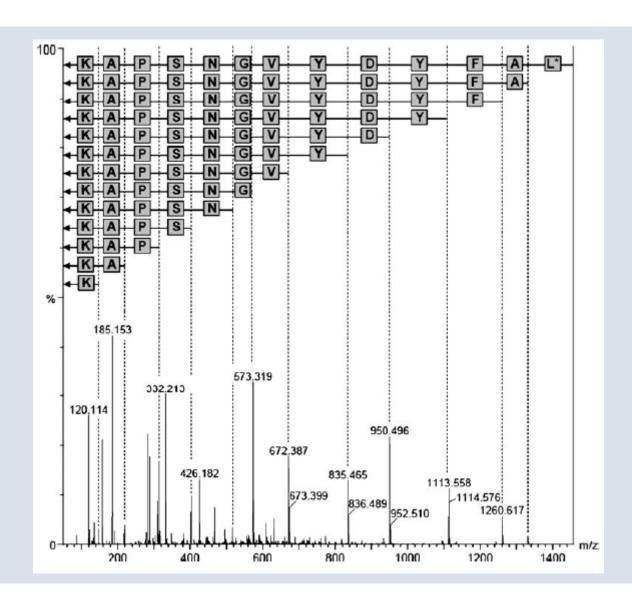
a,b,c - N-terminal side

x,y,z - C-terminal side

#### S-P-A-F-D-S-I-M-A-E-T-L-K (protonated mass 1410.6)

<u>mass</u> ⁺	<u>b-ions</u>	<u>y-ions</u>	mass+
88.1	S	PAFDSIMARTLK	1323.6
185.2	SP	AFD SIMAETLK	1226.4
256.3	SPA	FDSIMAETLK	1155.4
403.5	SPAF	DSIMAETLK	1008.2
518.5	SPAPD	SIMARTLK	893.1
605.6	SPAFDS	IMAETLK	806.0
718.8	SPAFDSI	MARTLK	692.3
850.0	SPAFDSIM	ARTLK	561.7
921.1	SPAFDSIMA -	ETPK	490.6
1050.2	SPAFDSIMAE	TLK	361.5
1151.3	SPAPDSIMART	TK	260.4
1264.4	SPAFDSIMAETL	K	147.2

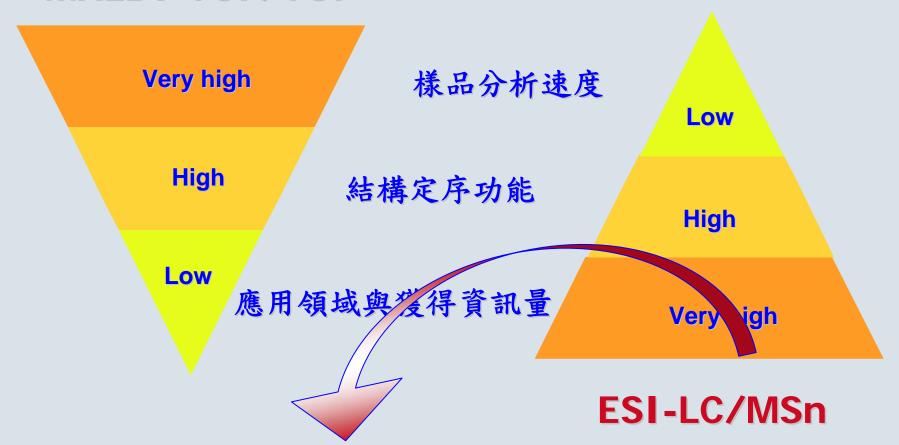




# LC/MSn vs. MALDI-TOF/TOF



#### MALDI-TOF/TOF



Metabolomics, Drug and Small Molecules

**Bruker Daltonics** 



# ESI-, MALDI-MS/MS Complementary

MKWVTFISLL LLFSSAYSRG VFRRDTHKSE IAHRFKDLGE EHFKGLVLIA FSQYLQQCPF DEHVKLVNEL TEFAKTCVAD ESHAGCEKSL HTLFGDELCK VASLRETYGD MADCCEKQEP ERNECFLSHK DDSPDLPKLK PDPNTLCDEF KADEKKFWGK YLYEIARRHP YFYAPELLYY ANKYNGVFQE CCQAEDKGAC LLPKIETMRE KVLASSARQR LRCASIQKFG ERALKAWSVA RLSQKFPKAE FVEVTKLVTD LTKVHKECCH GDLLECADDR ADLAKYICDN QDTISSKLKE CCDKPLLEKS HCIAEVEKDA IPENLPPLTA DFAEDKDVCK NYQEAKDAFL GSFLYEYSRR HPEYAVSVLL RLAKEYEATL EECCAKDDPH ACYSTVFDKL KHLVDEPONL IKONCDOFEK LGEYGFONAL IVRYTRKVPO VSTPTLVEVS RSLGKVGTRC CTKPESERMP CTEDYLSLIL NRLCVLHEKT PVSEKVTKCC TESLVNRRPC FSALTPDETY VPKAFDEKLF TFHADICTLP DTEKQIKKOT ALVELLKHKP KATEEOLKTV MENFVAFVDK CCAADDKEAC FAVEGPKLVV STQTALA **BSA** digest

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Pre-sequence
ESI
MALDI
both

Bruker Daltonics

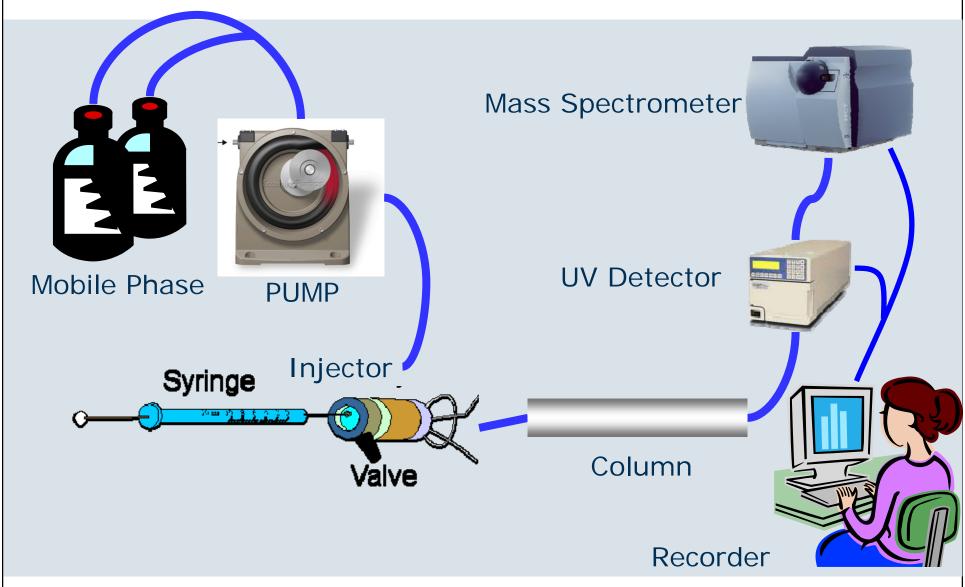


# What is "HPLC"?

High Performance Liquid Chromatography

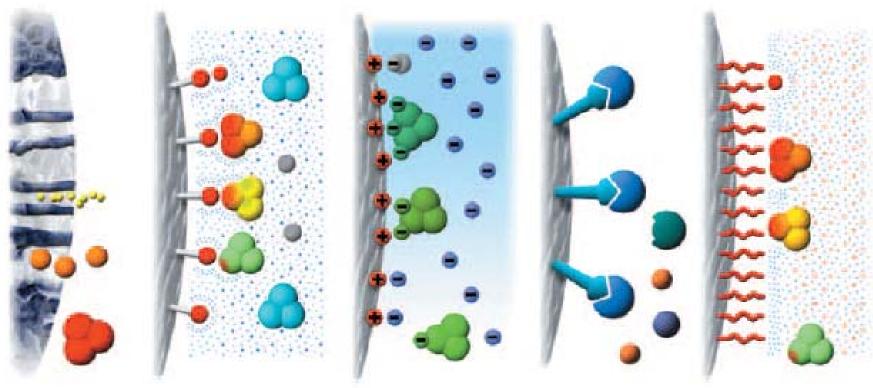
#### Individual Module of HPLC





## Category of Liquid Chromatography





**Gel Filtration** 

Ion Exchange

Reverse Phase

Hydrophobic Interaction

**Affinity** 

16 Bruker Daltonics

### Column for Reverse Phase Analysis





What should I know for a column selection?

# Chemistry of Reverse Phase Chromatography A. How much sample do you have?

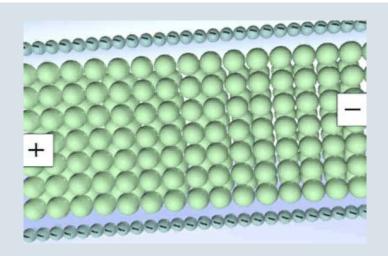


Column ID	Column Type	Flow Range (µL/min)	Optimum Flow per min	Analyte Capacity	Injector Volume
4.6 mm	Standard	500-3000	1000 ul	100ug~10ng	30 ul
2 mm	Microbore	100-1000	200 ul	10ug~1ng	5 ul
1 mm	Microbore	20-200	50 ul	1ug~100pg	1 ul
500 um	Microbore	5-50	12 ul	100ng~10pg	250 nl
300 um	Capillary	2-20	5 ul	10ng~1pg	128 nl
200 um	Capillary	1-10	2 ul	1ng~100fg	57 nl
100 um	Nanoscale	0.25-2.5	500 nl	100pg~10fg	14 nl
50 um	Nanoscale	0.05-0.5	100 nl	< 1pg	3 nl

Proteomics is here!!!

# Why Use Nanobore LC?





#### 平方倍數提高樣品分析靈敏度



Column	ID	Flow Rate	Relative [C]
Standard	4.6 mm	1 mL/min	1
Microbore	1 mm	50 μL/min	21
Capillary	320 µm	5 μL/min	206
Nanobore	75 μm	250 nL/min	3,750
Nanobore	50 μm	150 nL/min	8,450

### **Chemistry of Reverse Phase Chromatography**





#### **Chemistry of Reverse Phase Chromatography**



### Mobile Phase Composition



#### **Chemistry of Reverse Phase Chromatography**



### Mobile Phase Composition

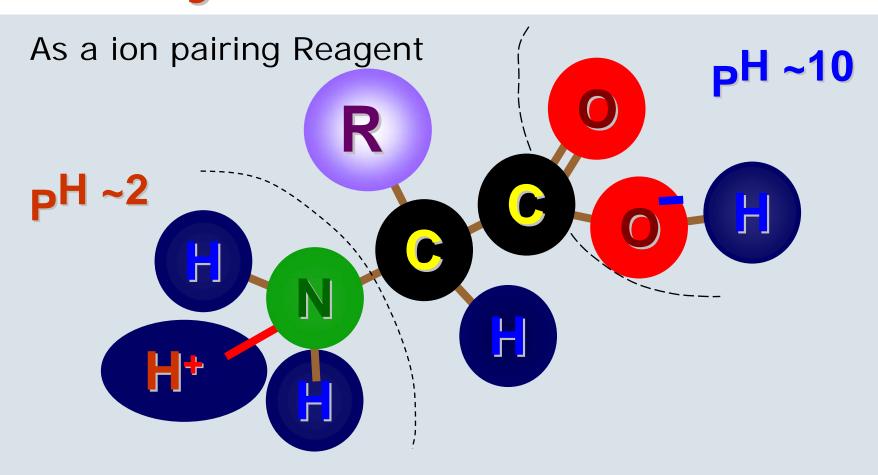
- A.  $H_2O$ , 0.1% FA (Formic Acid), pH: ~ 2
- B. CH<sub>3</sub>CN, 0.08% FA pH: ~2
  - \*. Filtered with 0.22um PVDF membrane

### Mobile Phase For Cation Exchange

20~50% ACN, no acid

# Why do we use FA?





# **Zwitterion Form**

### Why do we use these solvent?



TFA (Trifluoroacetic acid):

A. Improve chromatographic retention – Block residual active silanols

B. TFA is very effective at ion suppressing

Solution: Change to use FA (Formic acid).

Mobile Phase:

MeOH: Strong nonpolar solvent,

ACN: Less nonpolar effect, and good for peptide separation.

### 1D NanoLC Gradient Setup

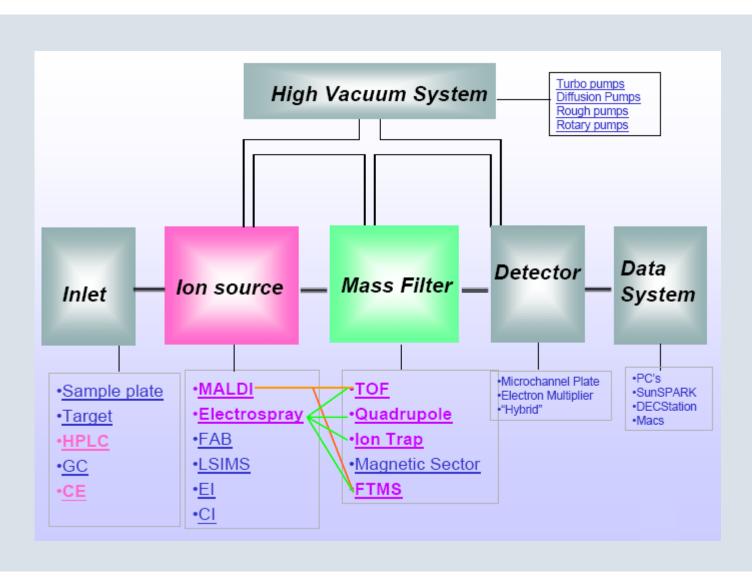


#### Common 1D LC Gradient

Time	A%	В%
0	97.0	3.0
2.0	97.0	3.0
2.1	92.0	8.0
26.1	76.0	24.0
31.1	61.0	39.0
33.0	5.0	95.0
38.0	5.0	95.0
38.5	97.0	3.0
52.0	97.0	3.0

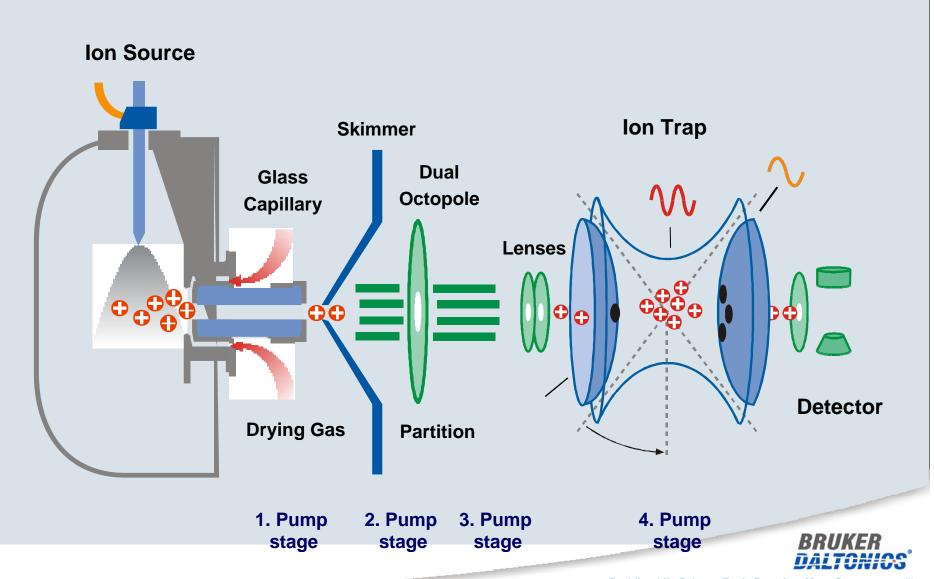
### Elements of Mass Spectrometer



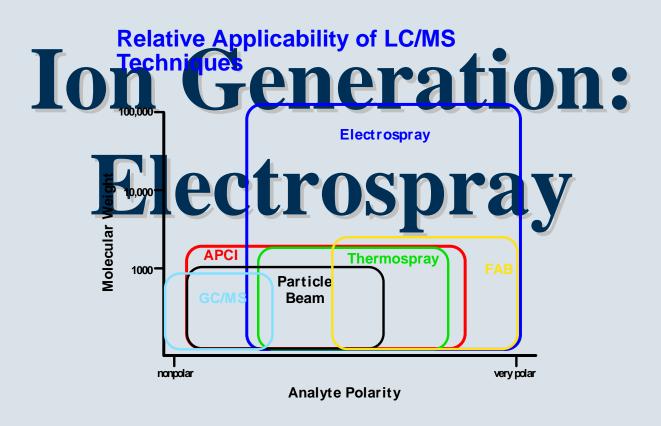


### **Experimental Setup of API - Ion Traps**







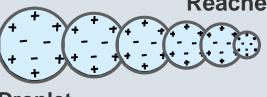


### **Generation of Ions (CRM)**



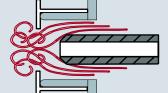
#### **Evaporation**

Rayleigh Limit Reached Coulomb Explosion







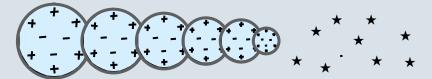


**Droplet** 

When the force of the Coulomb repulsion exceeds the surface tension of the droplet, the droplet explodes, producing charged daughter droplets that are subject to further evaporation.

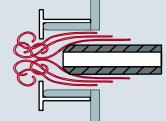
#### **Generation of Ions (IDM)**

#### **Evaporation**



# (Multiply Charged) Ion



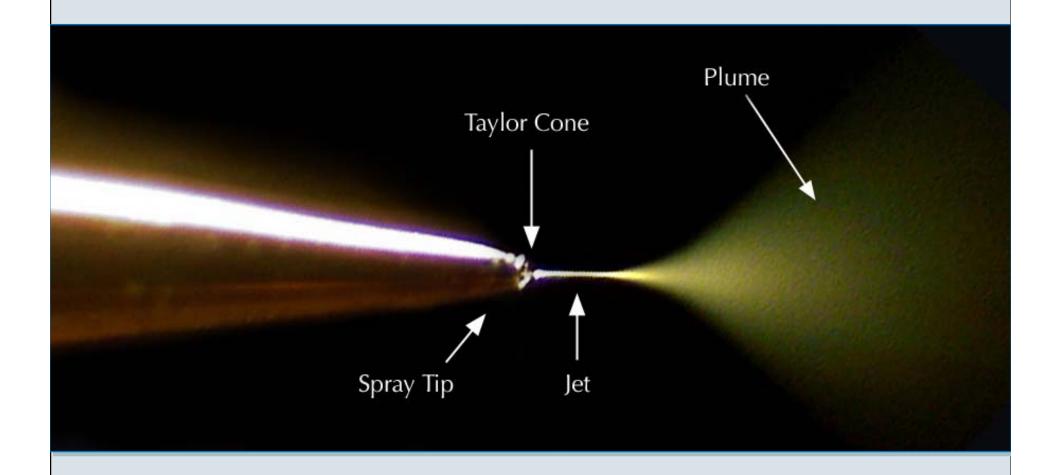


#### **Droplet**

As solvent evaporates from the droplet, the surface becomes highly charged. When the field created by the ions at the surface of the droplets exceeds the surface tension, ions are emitted directly into the gas phase.

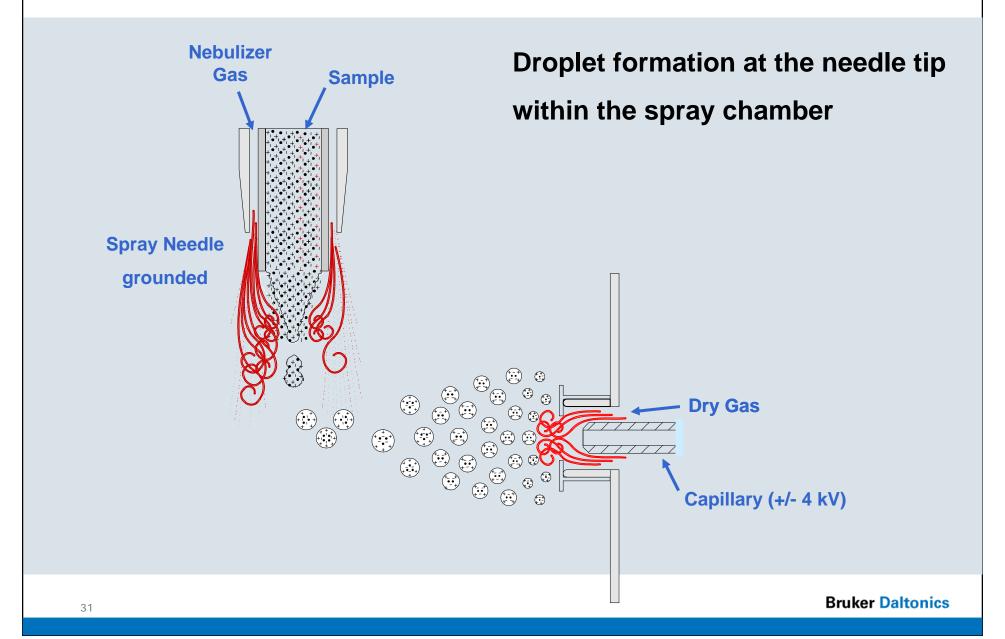






#### **Generation of Ions - Nebulization**

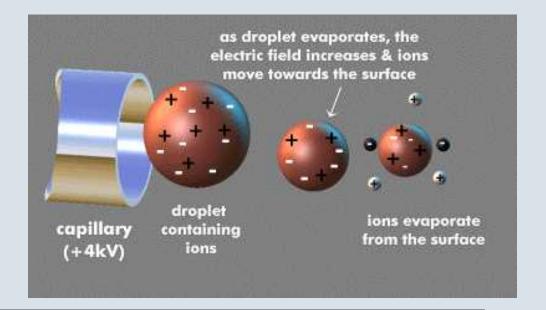






#### Ion generation for MS analysis

- Nebulization
- Desolvation
- Coulomb explosions
- Desorption



Under proper source conditions, individual ions only will enter the capillary. An unstable signal or capillary current may indicate a need for adjustment of gases, flow rate, or spray needle. See the User's Manual for Troubleshooting tips.



#### **Tune Parameters**

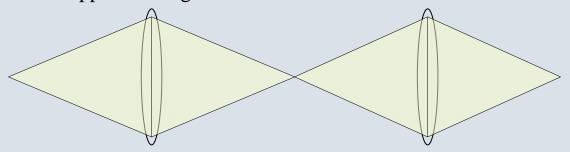


#### **Ion Optics**

The ion beam is continuous. Formed from the nebulizer spray and electrostatic fields, the ion beam is drawn into the capillary by the voltage exerted on it and by the pressure gradient. As it exits the capillary there is a natural tendency for the beam to spread out—to radiate like a beam of light.



The ion optics focus the ions into the analyzer and propel them against the pressure gradient. At each stage the ion beam wants to expand, so it has to be focused back down with the applied voltages.





#### **Tune Parameters**



#### **Ion Optics**

How the voltages affect the ions is largely mass dependent.

Smaller mass ions require less energy to control them.

and larger mass ions need more energy to direct them into the trap,







Thus we give the rule of thumb:

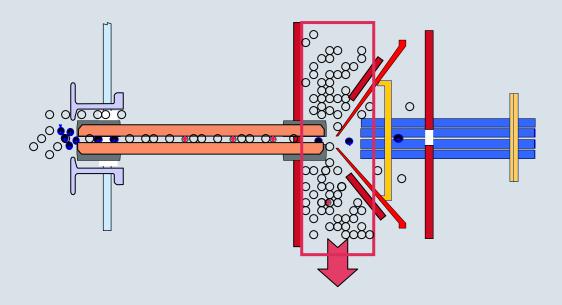
The larger the mass, the higher the value for the tuning parameters.







#### **Capillary Exit Voltage**



*In-source fragmentation*, or breaking the ions apart before they reach the ion trap, a balance must be struck between **Skimmer** and **Cap Exit** working in concert to set the voltage affecting the ions in this region. More unstable or fragile molecules need a lower Cap Exit voltage.



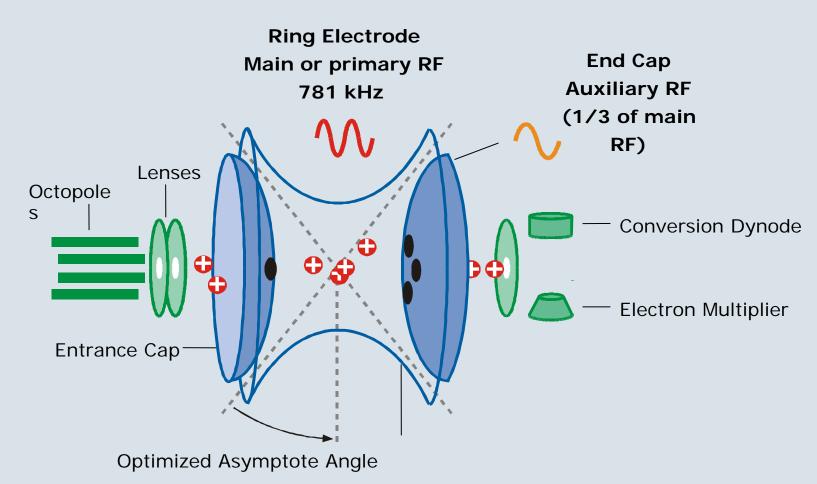


# Ion Trap

**THEORY** 

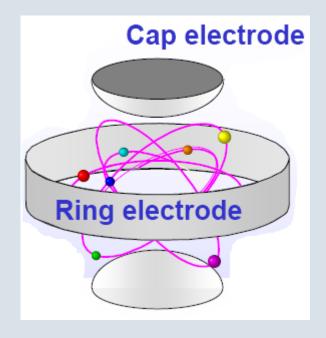
## **Ion Trap and Detector System**



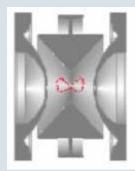


Ion Trap operations are performed by changing the voltages (amplitudes) of the primary and the auxiliary RF.







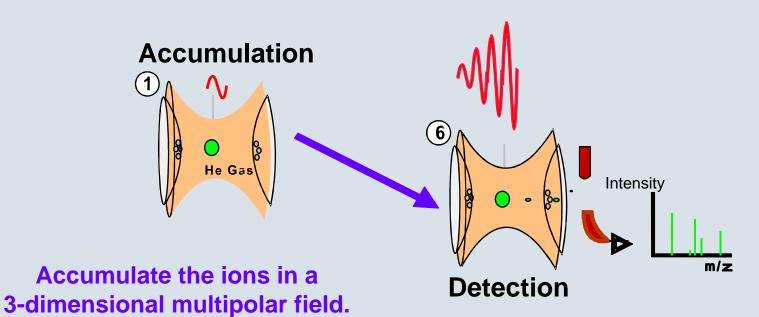








#### Generating a Mass Spectrum- overview

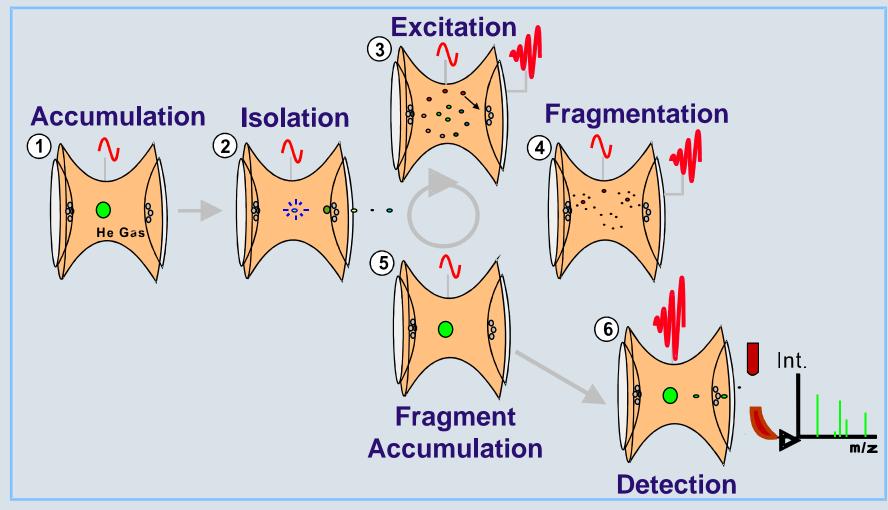


Scan the ions out of the trap in increasing mass by increasing the RF amplitude and measure the response at the detector.



## **Principles of Fragmentation in an Ion Trap**





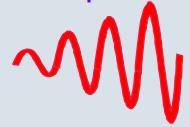
Generating MS<sup>n</sup> spectra

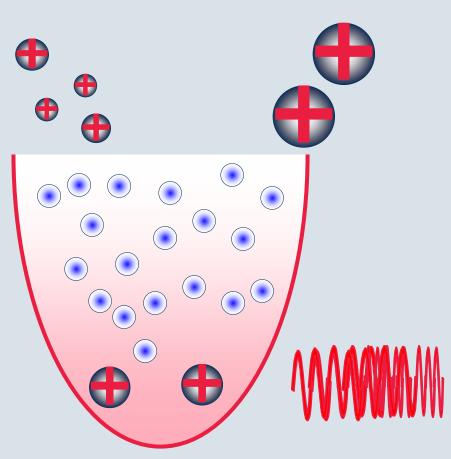
## Inside the trap - isolation



To isolate one small mass range, the low mass ions are ejected from the trap with a rapid scan.

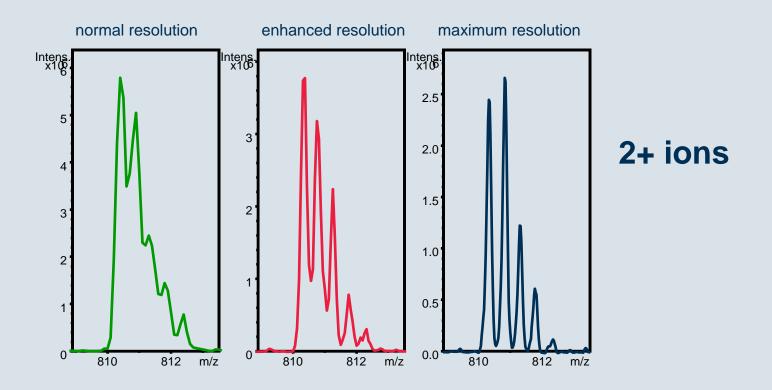
In addition, ions of a higher mass are ejected with a mix of frequencies.





#### Mass Resolution depends on the scan rate

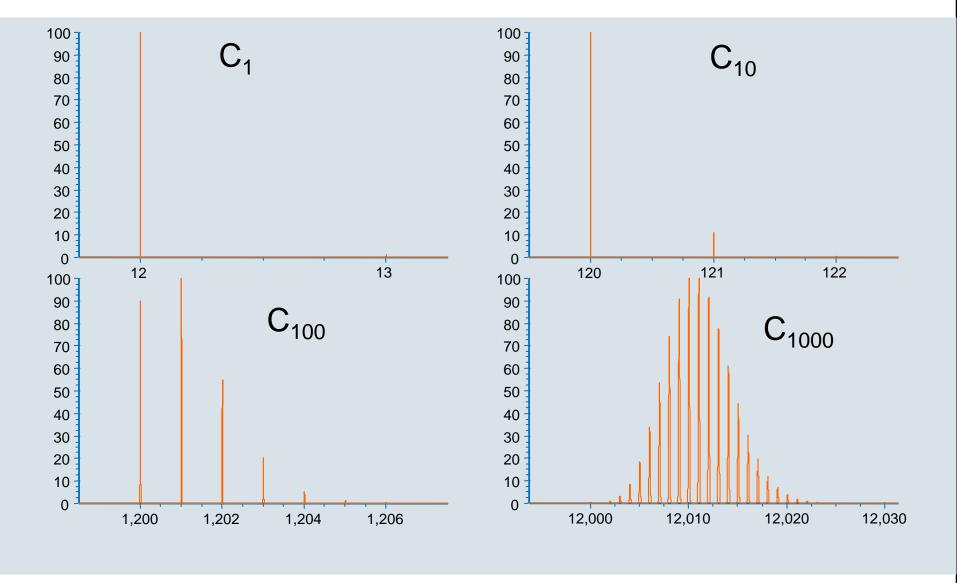




Here is an example of the resolution that results for a 2+ ion using different scan rates. All the scan rates are available over the full mass range of the trap. The trade off is that for more resolution there is a slower scan rate, so there are fewer data points across an LC peak. With infusion of a sample this time factor is not important.

#### **Isotopic Distribution Patterns**





#### Average & Monoisotopic Masses





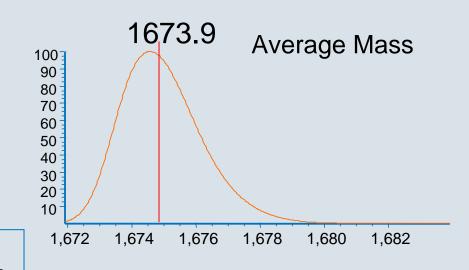
Res. 1'000

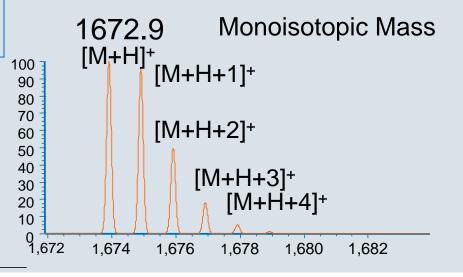
pQLYENKPRRPYIL MW 1672.9

Res. 10'000

Resolution = (m/z) / FWHM

What is the peak width at 10,000 resolution?

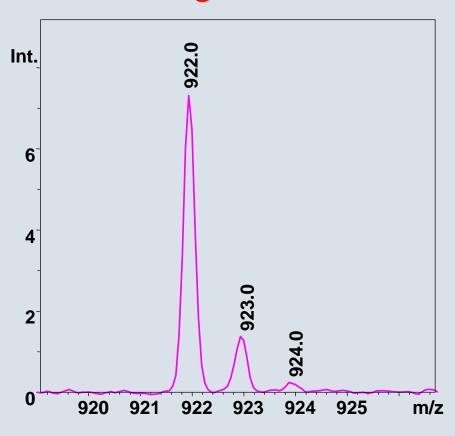


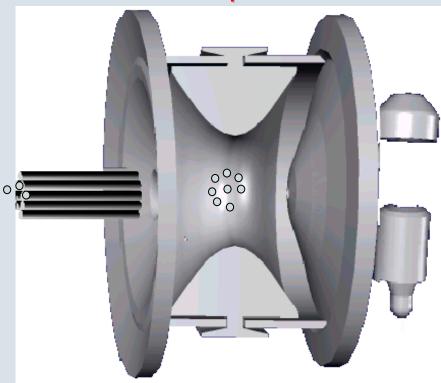


# Gating Ions:



#### Controlling the number of ions in the trap



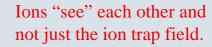


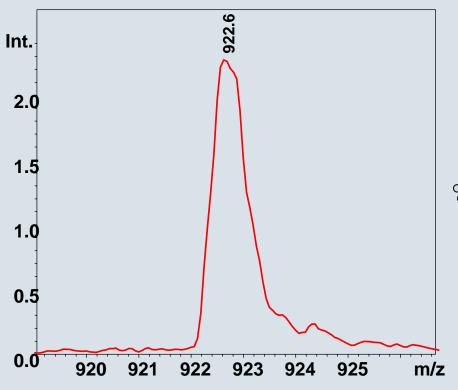
**Appropriate load gives good resolution** 

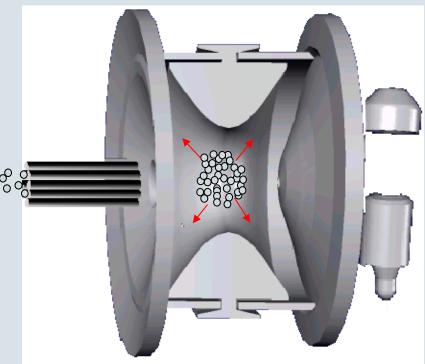
# Gating Ions:



#### Space Charging Effects





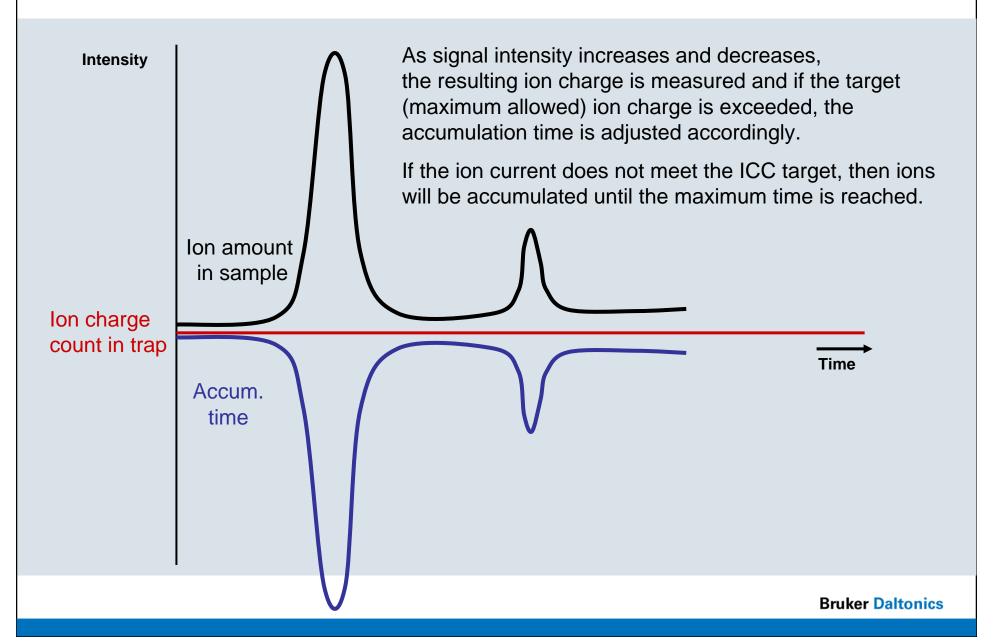


#### Too many ions: **OVERLOAD**!

- poor mass resolution
  - poor mass accuracy
  - poor peak area determination

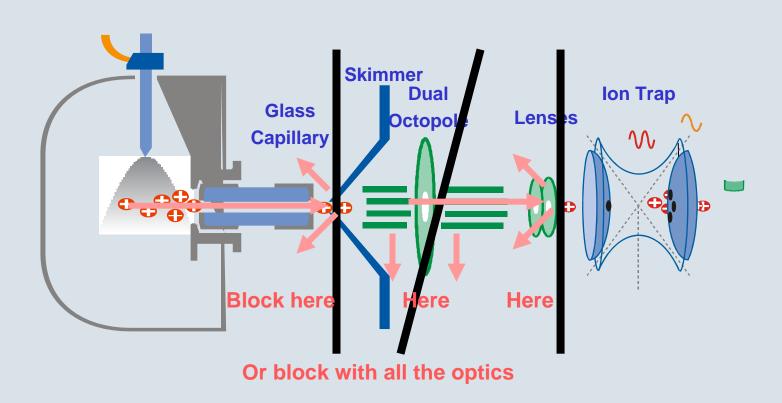
#### Gating ions: Ion Charge Control (ICC)





## **Blocking Voltages**





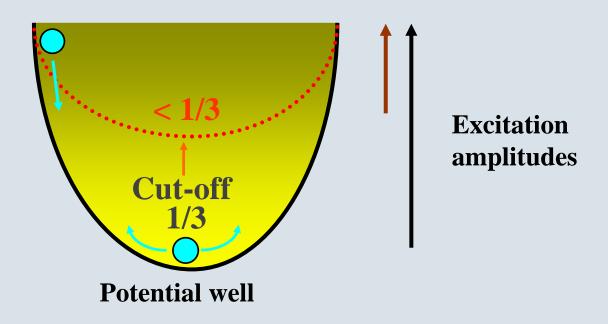
#### MS/MS below the 33% Limit



#### Changing the low m/z Cut-off for MS/MS Fragments

With MS/MS fragmentation, we use a preset cut-off of slightly under 1/3 of the mass that is isolated. This parameter is chosen to ensure that the isolated ion is excited most efficiently, to give the best MS/MS spectrum - the trade-off is that fragment ions of a mass below that 1/3 cut-off are not retained in the trap.

Other values can be set, to override the default cut-off. This is particularly useful if there is a fragment ion expected a little below the 1/4 value.



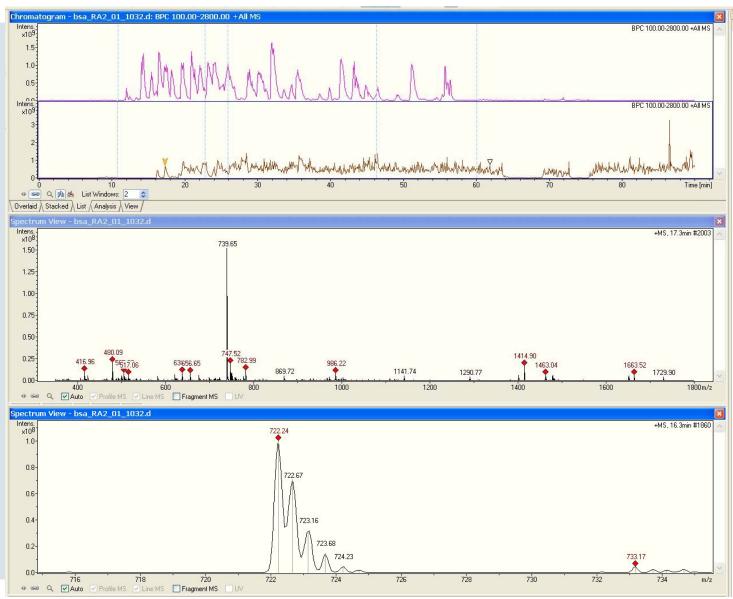




# 掃描取點越多越好!

- 1. How complex your sample is? →LC separation
- 2. Pick up valuable signals!  $\rightarrow$ 2+/3+ ions



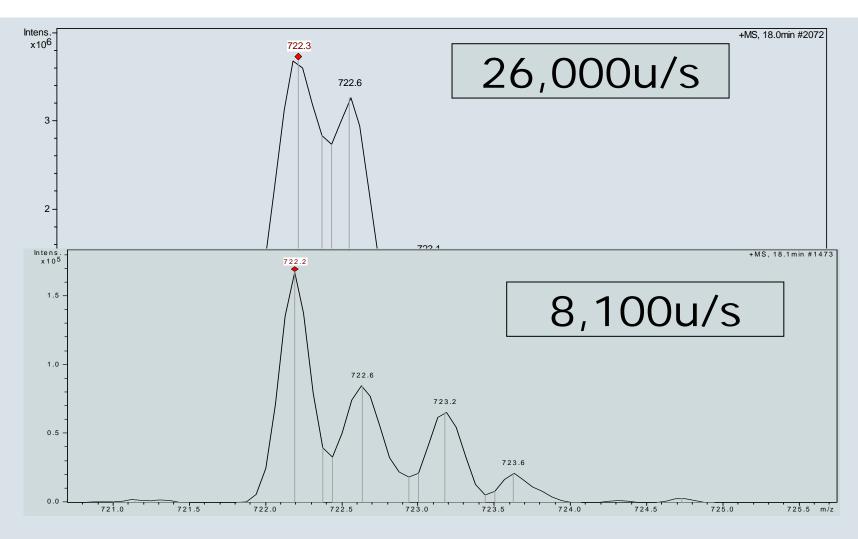




# 掃描速度越快越好?

速度越快,解析度越差 速度越慢,漏失訊號越多







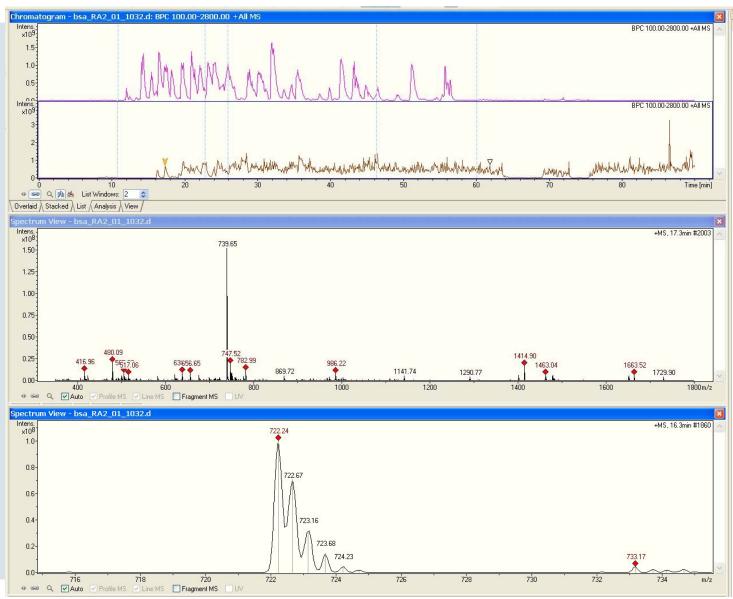
# 掃描範圍越大越好?

- 1. Ions charge state  $\rightarrow$  2+/3+
- 2.MS  $\leftarrow \rightarrow$  MS/MS setting
- 3. Duty cycle

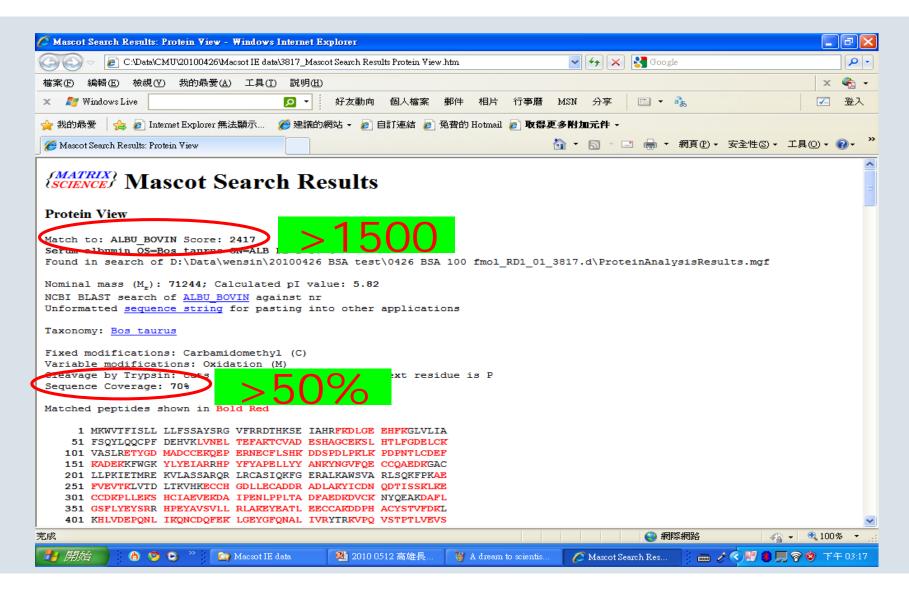


# Is the system good enough for sample test?

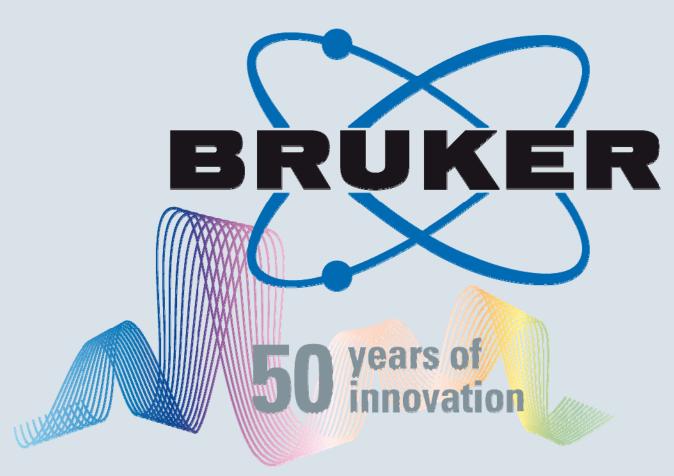












Bruker BioScience, Taiwan

2010/5/9